

Supplemental Material

Stochastic Ratchet Mechanisms for Replacement of Proteins Bound to DNA

S. Cocco, J.F. Marko, R. Monasson

I. TRANSITION MATRICES FOR THE DIFFERENT MODELS

We show in Table I the transition matrices W for the four models of Fig. 1 in the main text. States are numbered from 1 to $2N$ in the following way (the unbound state U is omitted as it is an absorbing state). The initially bound protein has N possible bound states T_i , where i is the number of attached units of the protein to the DNA. The 'invading' protein has, for each state of the bound protein, only two possible configurations: on or off ($j = 0, 1$), at the site corresponding to where the $(i + 1)$ th unit of the bound protein would be, which we call the 'zipping' site. There are thus $2N$ states, labelled by the index $S = 2i + j - 1$, for $i = 1, \dots, N$, $j = 0, 1$. The transition matrices W have therefore $2N$ lines and columns. Empty squares correspond to zero entries. For $N > 3$ the boundaries (two first and two last lines and columns) are the same, and the 2×2 central blocks are repeated $N - 2$ times on the band diagonal.

$$\begin{array}{c}
 W^{NZ} = \begin{array}{|c|c|c|c|c|c|} \hline -\rho-c-1 & \rho & \rho & \rho & & \\ \hline c & -2\rho & & & & \\ \hline 1 & & -\rho-c-1 & \rho & \rho & \rho \\ \hline & & c & -2\rho & & \\ \hline & & 1 & & -\rho-c & \rho \\ \hline & & & & c & -2\rho \\ \hline \end{array}, \quad
 W^{Z-NS} = \begin{array}{|c|c|c|c|c|c|} \hline -\rho-c-1 & \rho & \rho & & & \\ \hline c & -2\rho & & \rho & & \\ \hline 1 & & -\rho-c-1 & \rho & \rho & \\ \hline & & c & -2\rho & & \rho \\ \hline & & 1 & & -\rho-c & \rho \\ \hline & & & & c & -2\rho \\ \hline \end{array}, \\
 \\
 W^{Z-S-NSB} = \begin{array}{|c|c|c|c|c|c|} \hline -\rho-1 & & \rho & & & \\ \hline & -2\rho & & \rho & & \\ \hline 1 & & -\rho-1 & & \rho & \\ \hline & \rho & & -2\rho & & \rho \\ \hline & & 1 & & -\rho-c & \rho \\ \hline & & & & \rho & c \\ \hline & & & & & -2\rho \\ \hline \end{array}, \quad
 W^{Z-S-SB} = \begin{array}{|c|c|c|c|c|c|} \hline -\rho-1 & & \rho & & & \\ \hline & -2\rho & & \rho & & \\ \hline 1 & & -\rho-1-c & \rho & \rho & \\ \hline & \rho & c & -2\rho & & \\ \hline & & 1 & & -\rho & 1 \\ \hline & & & & & -1 \\ \hline \end{array}.
 \end{array}$$

TABLE I: Transition matrices W for the NZ (top, left), Z-NS (top, right), Z-S-NSB (bottom, left), and Z-S-SB (bottom, right) models with $N = 3$ binding units. All columns sum up to zero due to probability conservation, except the first two columns as states 1 (T_1) and 2 (R_1) may decay into the unbound state U, not represented in the matrices above.

II. UNBINDING RATES VS. CONCENTRATION FOR DIFFERENT ϵ AND N

In Fig. S1, Fig. S2, and Fig. S3 we show the unbinding rate r as a function of the concentration c for the Z-NS, Z-S-NSB and Z-S-SB models, for $\epsilon = 2, 3$ and different sizes N .

III. FIT OF MODEL PARAMETERS FROM EXPERIMENTAL DATA

To fit the Fis-Fis and CueR-CueR data we have proceeded as follows. We have first considered three possible elementary length $a = 0.5, 1$ and 2 nm. In Table II we give the elementary time t_0 and the unit of concentrations c_0 obtained in the three cases. The experiments give us access to the values of three quantities: the spontaneous dissociation rate $r(0)$, the order of magnitude of the concentration c_R at which replacement dominates over spontaneous dissociation, and the exchange rate R . We have first fitted the value N of the number of units for the Z-NS and Z-S models from the leading contribution to R (formulas given in main text), and we have then adjusted N and ϵ to fit the

a (nm)	0.5	1	5
t_0 (s)	$2 \cdot 10^{-10}$	$1.6 \cdot 10^{-9}$	$2 \cdot 10^{-7}$
c_0 (M)	8	1	0.008

TABLE II: Values of the time constant t_0 and of the unit of concentration c_0 for three different elementary lengths a .

spontaneous dissociation. The order of magnitude of c_R , given the fitted N and ϵ allows us to check the consistency of the model.

Interestingly we have found that the choice of a does not change the values, at the leading order, of R and c_R . Indeed at the leading order they both depend on the product $c_0 \times t_0$, which is independent of the value of a . More precisely, using the experimental values for R^{Fis} and R^{CueR} , we have found:

$$N_{Fis}^{Z-NS} \approx -\log_2(c_0 t_0 R_{Fis}) = 13.3, \quad (1)$$

and

$$N_{CueR}^{Z-S-NSB} \approx (c_0 t_0 R_{CueR})^{-1} - 1 = 21. \quad (2)$$

Given the approximate values of N above the concentration at which replacement start to dominates is given by

$$(c_R)_{Fis}^{Z-NS} \approx c_0 t_0 r_{Fis}(0) 2^N \approx 1 \text{ nM}, \quad (3)$$

and

$$(c_R)_{CueR}^{Z-S-NSB} \approx c_0 t_0 r_{CueR}(0) (N + 1) \approx 17 \text{ nM}. \quad (4)$$

Then we have used the spontaneous dissociation rate $r(0)$ to estimate, given N , the order of magnitude of the binding energy ϵ through

$$\epsilon \approx -\log(t_0 r(0))/N, \quad (5)$$

for both Fis and CueR data. Note that the Z-NS model is not compatible with CueR data because we would obtain $N_{CueR}^{Z-NS} = 4$, which would corresponds to large binding energies of $\epsilon = 5.7, 5.2, 4 k_B T$, for, respectively, $a = 0.5, 1, 5$ nm. Conversely, Z-S-NSB is not compatible with Fis data because we would obtain $N_{Fis}^{Z-S-NSB} = 10,000$.

We have then tuned the value of $N_{CueR}^{Z-S-NSB}$ and N_{Fis}^{Z-NS} to precisely fit R when the whole expression of the exchange rate is taken into account (see Eqs. (8) and (9) of main text), in particular the multiplicative factors in $1 - \rho = 1 - e^{-\epsilon}$. The final value for $N_{CueR}^{Z-S-NSB}$ ranges from 13 to 16 depending on the value of a (see Fig. 3 of the main text). There is less variability in the value of N_{Fis}^{Z-NS} , due to the exponential dependence of R upon N . The best fit is obtained in the three cases for $N = 14$; the best value for a is $a = 5$ nm. Finally, once N is fixed, a fine tuning of ϵ is easily done to reproduce the spontaneous dissociation rate.

We show in Fig. S4 the fits of the Fis-Fis, Fis-Hu, and CueR-CueR experiments presented and cited in the main text. In each panel three fits are presented, corresponding to the three values of the elementary length $a = 0.5, 1, 5$ nm. The values of N and ϵ are given in the panels. The slope of the curves are the exchange constants R :

- The experimental Fis-Fis exchange constant $R_{Fis-Fis} = 6 \cdot 10^4 \pm 3 \cdot 10^3 \text{ M}^{-1} \text{ s}^{-1}$ has to be compared with $R_{Fis-Fis}^{Z-NS} = 4.9 \cdot 10^4, 5.1 \cdot 10^4, 6 \cdot 10^4 \text{ M}^{-1} \text{ s}^{-1}$ for, respectively, $a = 0.5, 1, 5$ nm. The replacement concentration is $(c_R)_{Fis}^{Z-NS} = 2, 2, 1.6 \text{ nM}$ for $a = 0.5, 1, 5$ nm.
- The experimental Fis-Hu exchange constant $R_{Fis-Hu} = 2.7 \cdot 10^3 \pm 5 \cdot 10^2 \text{ M}^{-1} \text{ s}^{-1}$ has to be compared with $R_{Fis-Hu}^{Z-NS} = 2.3 \cdot 10^3, 2.6 \cdot 10^3, 4 \cdot 10^3 \text{ M}^{-1} \text{ s}^{-1}$ for, respectively, $a = 0.5, 1, 5$ nm. The replacement concentration is $(c_R)_{Fis-Hu}^{Z-NS} = 400, 370, 180 \text{ nM}$ for $a = 0.5, 1, 5$ nm.
- The experimental CueR-CueR exchange constant is $R_{CueR-CueR} = 2.8 \cdot 10^7 \text{ M}^{-1} \text{ s}^{-1}$ has to be compared with $R_{CueR-CueR}^{Z-S-NSB} = 2.8 \cdot 10^7, 2.9 \cdot 10^7, 3.1 \cdot 10^7 \text{ M}^{-1} \text{ s}^{-1}$ for, respectively, $a = 0.5, 1, 5$ nm. We obtain the replacement concentration $(c_R)_{CueR-CueR} = 19, 16, 17 \text{ nM}$ for $a = 0.5, 1, 5$ nm.

IV. SCALING OF THE REPLACEMENT RATE R WITH N

We start by writing the average unbinding time as

$$t_{unb}(c) = (1 - P_R) t_{unb}(0) + P_R t_{unb-R} , \quad (6)$$

where P_R is the probability that unbinding occurs (at least partially) through the replacement pathway, $t_{unb}(0) = 1/r(0)$ is the average unbinding time through the thermal pathway only, and t_{unb-R} is the average unbinding time through the replacement pathway.

In the limit of small solution-phase protein concentrations, *i.e.* for small rates c , we expect P_R to scale linearly with c , $P_R \simeq p_R c$, and, thus, that $P_R \ll 1$. In addition, as unbinding through replacement is much faster than through thermal activation alone, we have $t_{unb-R} \ll t_{unb}(0)$. Taking the inverse of (6) we obtain

$$r(c) = \frac{1}{t_{unb}(c)} \simeq r(0) + p_R c r(0) . \quad (7)$$

Comparing with Eq. (3) in the main text we see that the replacement rate R coincides with

$$R = p_R r(0) . \quad (8)$$

To calculate p_R , which corresponds to the linear term in the expansion of P_R in powers of c , we consider 'replacement' paths with only one binding event for the invader. The scaling behavior of p_R with N can be simply guessed from the most likely scenario leading to dissociation through the replacement pathway, see main text and blue configurations sketched in Fig. 1 therein. Below, we consider all contributions to p_R to find back the exact expression for R given in Eqs. (7), (8) and (9) in the main text for the three Z- kinetic models; the unbinding time for the NZ model at small c is not significantly larger than the thermal rate, and the asymptotic dependence of the replacement rate with N is easy to determine, see main text.

We will use the following result, true for the thermal pathway alone ($c = 0$): the average time spent in state T_i (before the protein eventually unbinds) is

$$\tau_i = (1 - \rho) \rho^{N-i} (1 + O(\rho^N)) / r(0) , \quad (9)$$

where $r(0) = (1 - \rho)^2 \rho^N$ to the leading order in powers of ρ , see main text. The sum of τ_i over all states $i = 1, \dots, N$ coincides with our expression for the thermal unbinding time, $t_{unb}(0) = 1/r(0)$.

A. Z-NS model

One possible scenario of unbinding due to replacement, considered in the main text, consists in reaching state R_N from state T_N . As the average time spent in state T_N is $\tau_N = (1 - \rho)/r(0)$ the probability that the invader eventually binds is, to the lowest order in c , $\simeq c \tau_N$. Next, the probability to stay and go all the way up along the alternative replacement pathway is 2^{-N} , because each transition $R_i \rightarrow R_{i-1}$ has the same rate as the transition $R_i \rightarrow T_i$ (invader detachment). The contribution of this scenario to the replacement rate is

$$\tau_N 2^{-N} r(0) = (1 - \rho) 2^{-N} \quad (10)$$

Other replacement scenarios, with a single invader-binding event, are as follows: the system spends time in the thermal pathway, undergoes the transition $T_k \rightarrow R_k$ at some level (number of bound units) k , remains in the replacement pathway until level $\ell (\leq k)$, and goes back to the thermal pathway through $T_\ell \rightarrow R_\ell$, until thermal binding occurs (the scenario above corresponds to the case $k = N$, without back transition to the thermal pathway). The probability of climbing (without leaving) the replacement pathway from T_k to T_ℓ is $2^{-(k-\ell)}$.

We then need to calculate the probability q_ℓ that the system reaches back the thermal pathway in state T_ℓ . Let us start by estimating the probability μ_ℓ that, once in state ℓ , the protein will unbind quickly, *i.e.* in a time much lower than the average time t_{unb} in the thermal pathway. The following physical picture is useful to compute μ_ℓ . A particle is undergoing a biased random walk on discrete sites $i \geq 0$ along the 1D semi-infinite line; $i = 0$ is an absorbing site. The random walk is biased by a force F pushing the particle towards the right (large i), with $F = -\log \rho$. μ_ℓ is the probability that, starting from state $i = \ell$, the particle will reach the absorbing state, rather than being attracted towards $i \rightarrow \infty$. We obviously have

$$\mu_\ell = \frac{1}{1 + \rho} \mu_{\ell+1} + \frac{\rho}{1 + \rho} \mu_{\ell-1} , \quad \forall \ell = 2, \dots, N , \quad (11)$$

with the boundary conditions $\mu_0 = 1$ and $\mu_\infty = 0$, which gives

$$\mu_\ell = \rho^\ell, \quad (12)$$

for all $\ell \geq 0$. Formally speaking, μ_ℓ is the probability that the system reaches the U state in a finite time (when $N \rightarrow \infty$), conditioned to the starting state, T_ℓ . The probability q_ℓ that the system reaches back the thermal pathway in state T_ℓ conditioned to fast unbinding is, according to Bayes' rule,

$$q_\ell = \frac{\mu_\ell}{\sum_{i \geq 0} \mu_i} = (1 - \rho) \rho^\ell. \quad (13)$$

The sum of the contributions corresponding to all pairs (k, ℓ) gives the replacement rate

$$R^{Z-NS} = \sum_{\ell \leq k \leq N} \tau_k 2^{-(\ell-k)} q_\ell r(0) = (1 - \rho)^2 \sum_{\ell \leq k \leq N} \rho^{N-k+\ell} 2^{-(k-\ell)} \quad (14)$$

$$= (1 - \rho)^2 \sum_{0 \leq m \leq N} (N + 1 - m) \rho^{N-m} 2^{-m} = \frac{(1 - \rho)^2}{(1 - 2\rho)^2 2^N}, \quad (15)$$

to the leading order in N , in agreement with Eq. (7) in the main text.

B. Z-S-NSB model

In the Z-S-NSB model the alternative scenario to thermal unbinding starts with a transition $T_N \rightarrow R_N$. Later the system changes states in the replacement pathway, until unbinding occurs. We need to estimate the probability Q that this scenario occurs, rather than the system reaches back state T_N from R_N , and unbinding occurs through the thermal pathway. Let $\mu_{i,N}$, with $1 \leq i \leq N$ the probability that the system never leaves the replacement pathway (and unbinding eventually occurs), starting from state T_i ; we want to calculate $Q = \mu_{N,N}$. According to Fig. 1 in the main text (bottom & left panel), the rates of the forward ($R_i \rightarrow R_{i-1}$) and backward ($R_i \rightarrow R_{i+1}$) transitions are identical (away from the boundaries). Hence the $\mu_{i,N}$ s fulfill the recurrence equations

$$\mu_{i,N} = \frac{1}{2}(\mu_{i-1,N} + \mu_{i+1,N}), \quad \forall i = 2, \dots, N. \quad (16)$$

The above equation can be extended to the states $i = N$ and $i = 1$, with the prescriptions $\mu_{N+1,N} \equiv 0$ (which expresses the fact that the system cannot re-enter the replacement pathway after having left it), and $\mu_{0,N} \equiv 1$ (where the subscript 0 stand for the unbound state U here). The solution to these equations is

$$\mu_{i,N} = \frac{N + 1 - i}{N + 1}. \quad (17)$$

We deduce that the replacement rate is given by

$$R^{Z-S-NSB} = \tau_N \mu_{N,N} r(0) = \frac{1 - \rho}{N + 1}, \quad (18)$$

in perfect agreement with Eq. (8) in the main text.

C. Z-S-SB model

The replacement rate for the Z-S-SB model is calculated in the same way as for the Z-S-NSB model. The only differences are: the replacement pathway is entered through the $T_{N-1} \rightarrow R_{N-1}$ transition, and the 'length' (number of states) of this replacement pathway is now $N - 1$ instead of N . We readily obtain, using (9) and (17),

$$R^{Z-S-SB} = \tau_{N-1} \mu_{N-1,N-1} r(0) = \frac{\rho(1 - \rho)}{N}, \quad (19)$$

in agreement with Eq. (9) in the main text.

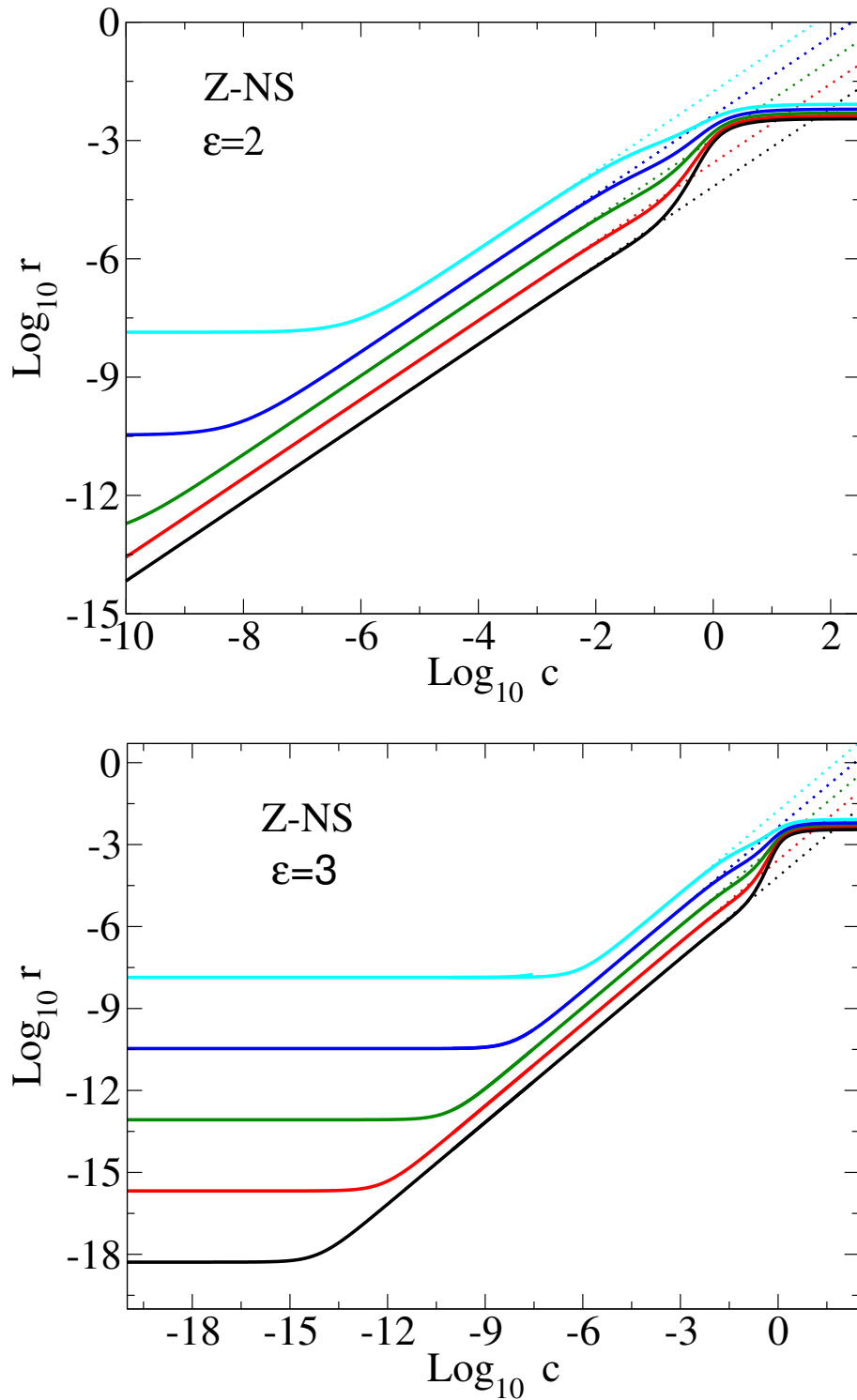


FIG. S1: Unbinding rate as a function of the concentration for the Z-NS model, and for different protein sizes N ranging from 6 to 14, and monomer binding energy $\epsilon = 2$ and 3 (in units of $k_B T$) in log-log scale. Full lines: numerical calculation of the unbinding rate. Dotted lines: linear approximations to the unbinding rate, see main text.

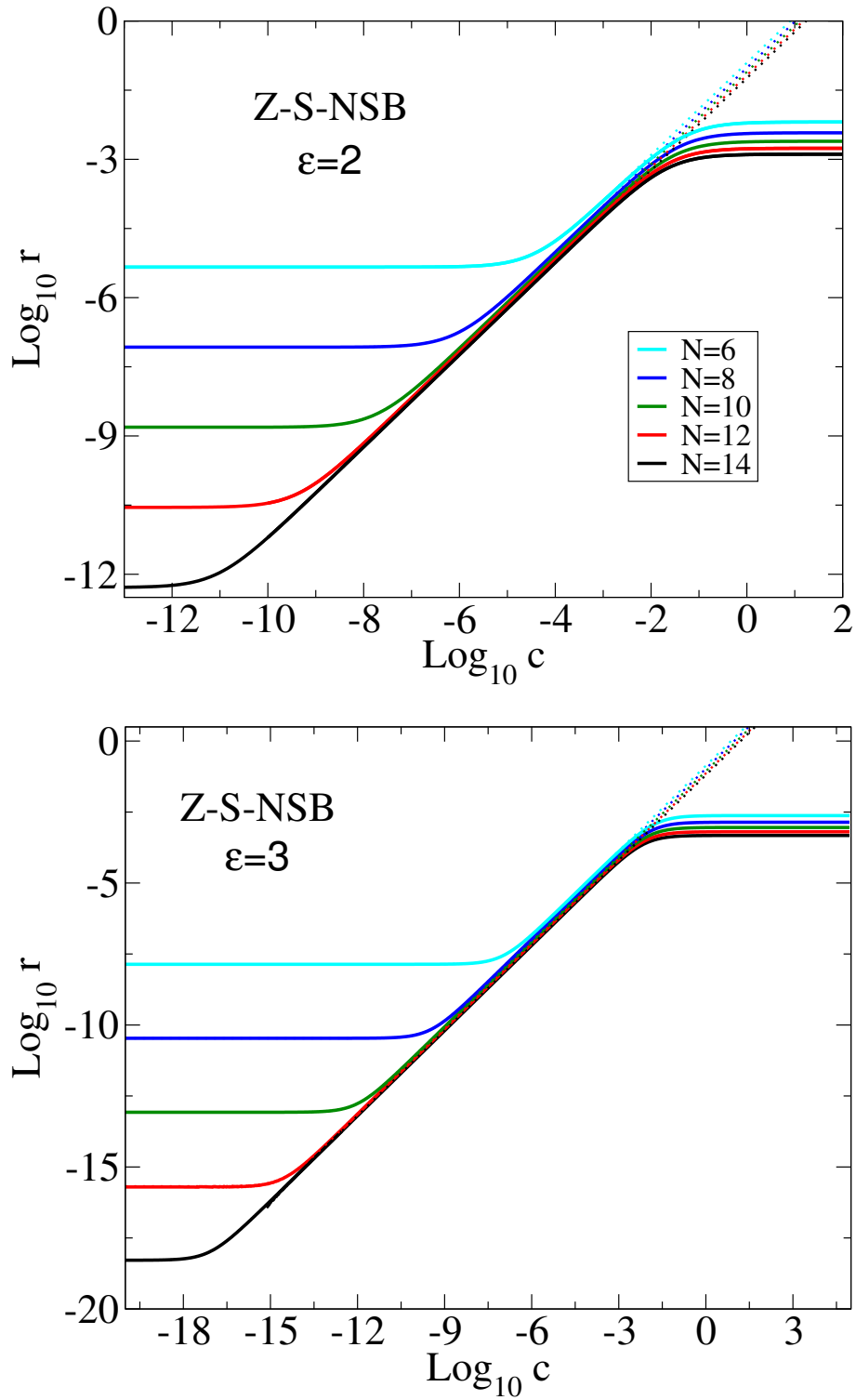


FIG. S2: Unbinding rate as a function of the concentration for the Z-S-NSB model, and for different protein sizes N ranging from 6 to 14, and monomer binding energy $\epsilon = 2$ and 3 (in units of $k_B T$) in log-log scale. Full lines: numerical calculation of the unbinding rate. Dotted lines: linear approximations to the unbinding rate, see main text.

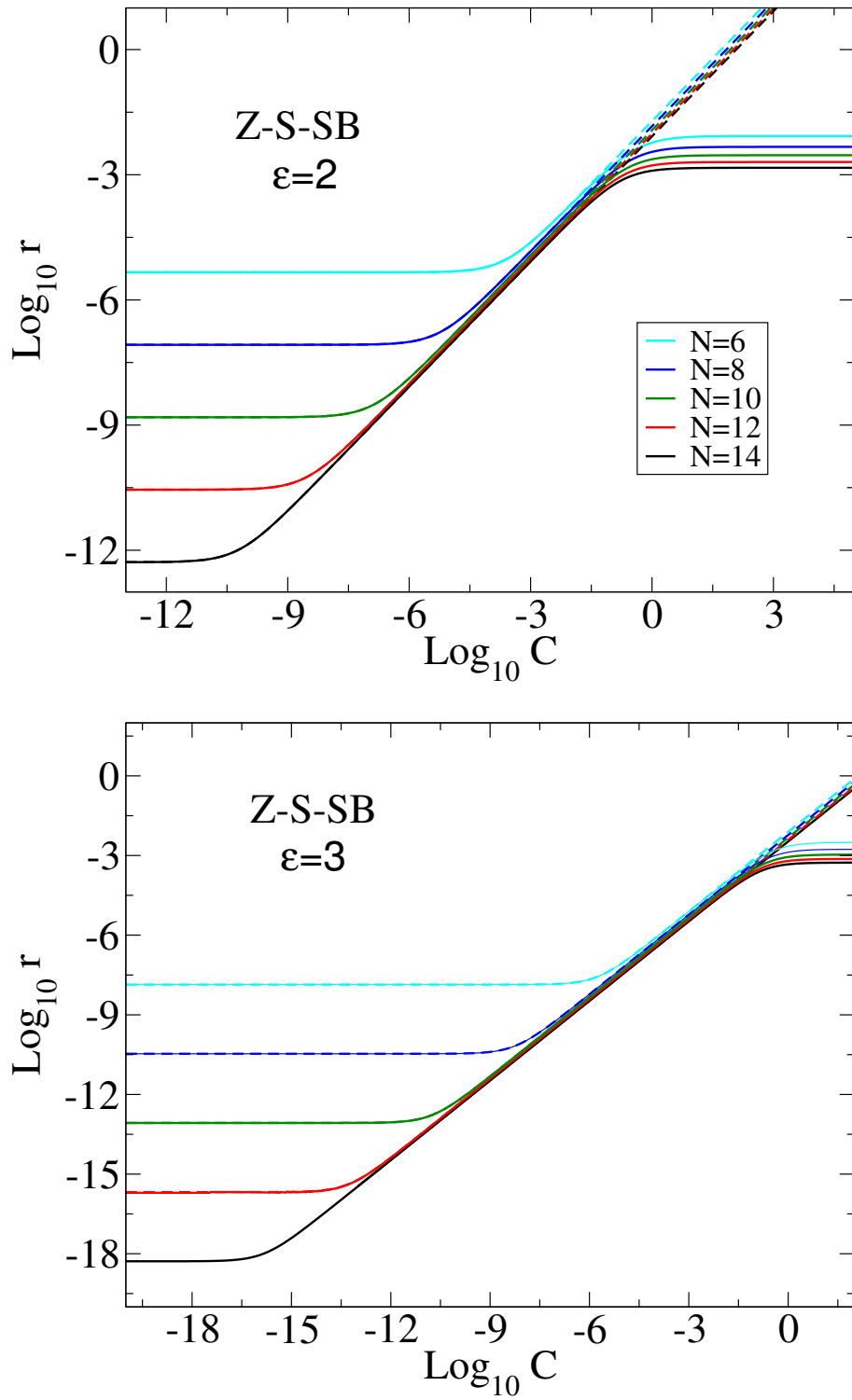


FIG. S3: Unbinding rate as a function of the concentration for the Z-S-SB model, and for different protein sizes N ranging from 6 to 14, and monomer binding energy $\epsilon = 2$ and 3 (in units of $k_B T$) in log-log scale. Full lines: numerical calculation of the unbinding rate. Dotted lines: linear approximations to the unbinding rate, see main text.

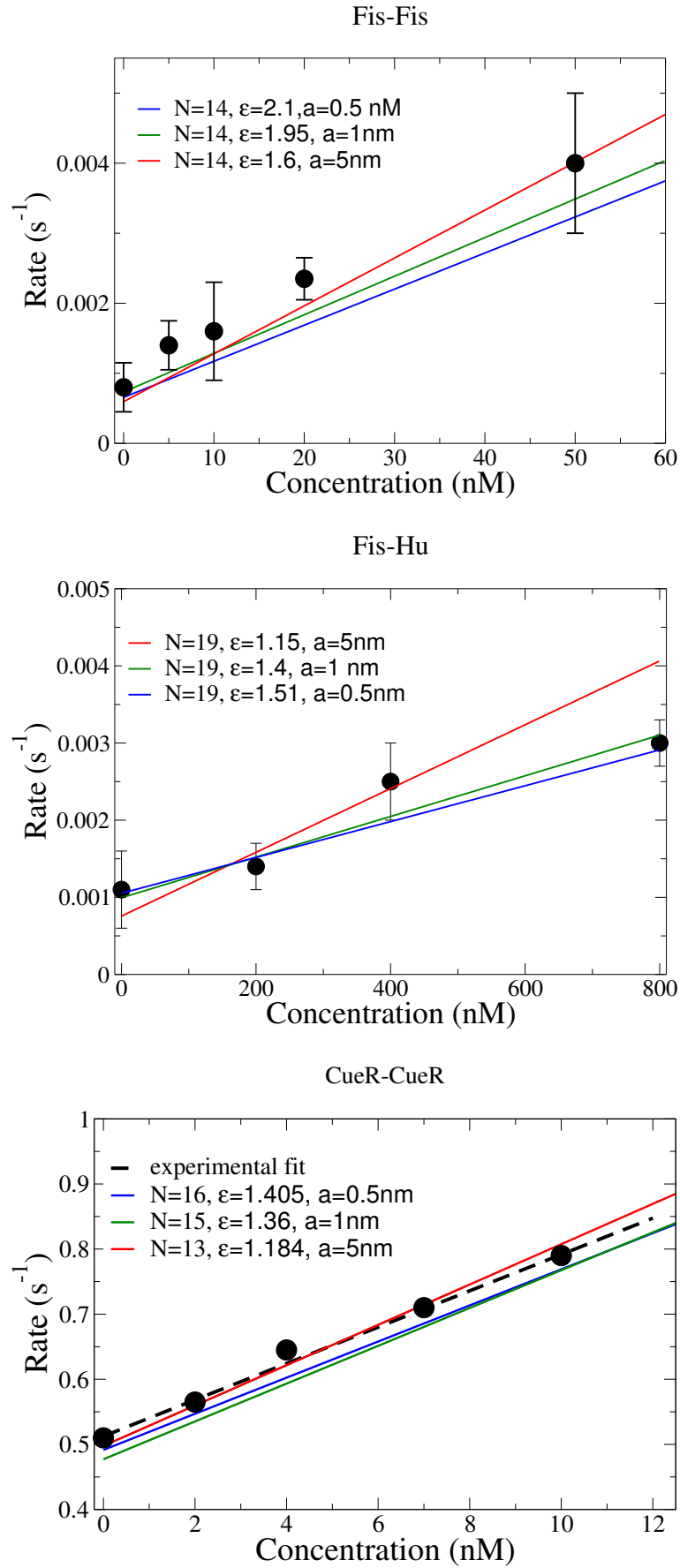


FIG. S4: Fit of the experimental data on concentration dependent dissociation rates of Fis proteins bounded on DNA as a function of concentration of Fis proteins (top) and Hu proteins (middle) in solution with the Z-NS model. Bottom: fit of CueR dissociation rates as a function of CueR concentration in solution with the Z-S-NSB model. See text for details on the parameter values.